

Isoproturon (IPU) Electrochemical Sensing Based on a Polymethylmethacrylate Ferrite (PMMA/M(FexOy)) Nanocomposite Modified Electrode

Sameena Mehtab*, M. G. H. Zaidi, Priya Bhatt,
Pragati Joshi and Tithi Agarwal

*Department of Chemistry, College of Basic Science and Humanities,
G. B. Pant University of Agriculture and Technology,
Pantnagar, Uttarakhand, India*

*Corresponding author. E-mail address: smiitr@gmail.com

Received 27/03/20; accepted 10/08/2021

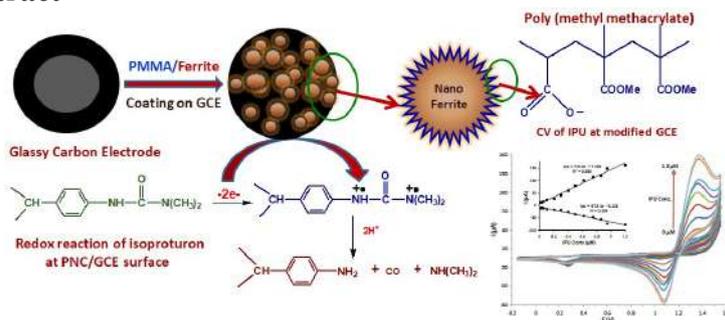
<https://doi.org/10.4152/pea.2022400305>

Abstract

The use of pesticides has been increased in recent years, to enhance crops productivity, which may lead to a serious global concern of environmental pesticides monitoring. Isoproturon (IPU: 3-(4-isopropylphenyl)-1,1-dimethylurea) is an herbicide widely used in wheat crops. However, when it enters in the ecosystem, it is heavily toxic to humans. Thus, there is an urgent need to develop sensitive and selective IPU detection methods. In the present work, a novel polymethylmethacrylate/ferrite (PMMA/M(FexOy)) polymer nanocomposite (PNC) modified glassy carbon electrode (GCE) was developed for IPU detection. The PNC/GCE surface morphology was characterized by scanning electron microscopy (SEM) and atomic force microscopy (AFM). PNC electrochemical characterization (EC) was performed by cyclic voltammetry (CV) that showed a quasi-reversible redox behavior. PNC/GCE demonstrated an excellent square wave voltammetric (SWV) response towards IPU, with a limit of quantification (LOQ) of 1.98×10^{-7} M, and a limit of detection (LOD) of 6.5×10^{-8} M, in 1 M HClO₄ (perchloric acid), at pH 2.0. EC investigations reflected a peak current that was linearly related to IPU concentrations, with a high detection sensitivity. It also showed much better CV and SWV IPU responses than those of a bare GCE, and further environmental stability, without a high influence of common interfering ions.

Keywords: IPU, PPMA/M(FexOy), EC, CV and SWV.

Graphical abstract



Introduction

Pesticides have greatly contributed to a global increase in food productivity, profits for farmers and diseases prevention. Pesticides do not always stay in the applied location, as they are mobile in the environment, and often move through water, air and soil. Pesticides large scale application of has resulted in their remarkable persistence in the environment, representing a threat for the entire ecosystem [1]. Conventional methods employed for pesticides detection are gas chromatography and high performance liquid chromatography (HPLC) [2, 3]. Non-chromatographic methods, such as UV-Vis, infrared spectroscopy, solid-phase extraction, enzymatic techniques and mass spectrometry have also been reported for pesticides detection [4, 5]. These techniques for pesticides analysis are highly expensive, and not completely acceptable for the routine monitoring of a large number of samples. Consequently, rapid and reasonable methods of analysis are required to complement these techniques. EC methods are a key alternative to these complicated techniques, due to their extremely high sensitivity, selectivity and simplicity in experimental procedures [6-8]. Pesticides detection has been achieved through many EC techniques, such as CV [9], Differential Pulse Voltammetry (DPV) [10] and SWV [11]. SWV has the potential of monitoring pesticides through the use of GCE, by minimizing the charging current, which results in a sharp electrochemical response towards the analyte, due to an intense faradic current [12]. Over past decades, to achieve a high electrochemical response on pesticides detection, GCE surfaces have been modified by different electroactive materials *viz.*, metal oxides [13-15], clays [16], carbonaceous nanomaterials [17-20], PNC with metal oxides [21], metals [22-23] and conducting polymers [24].

IPU is a highly selective and systematic phenyl urea herbicide, chiefly effective against innumerable annual grasses and several weeds [25]. It can easily enter in a human food chain by migration through crops in soil and water. Due to a slow degradation, its concentration goes on increasing in both ground and drinking water [26]. The maximum acceptable IPU concentration in drinking water, set by the European Union, is 0.1 g/L [27]. The continuous use of IPU over many years has resulted in the development of resistance in *Phalaris Minor*, causing huge losses in wheat productivity [28]. IPU use can induce oxidative stress and reduction in the chlorophyll content in wheat plants [29]. Its toxic effects have also been reported in the reproductive system of albino rats [30] and aquatic organisms, such as diatoms [31]. Some IPU cytogenetic effects have also been seen in human lymphocytes [32]. There is a vital requirement for the establishment of a fast, reliable and economical analytical technique for IPU determination.

In recent times, a lot of attention has been given to the synthesis and employment of the PNC polymer in the detection of several pesticides through EC [33-35]. It is reported in literature that IPU can be detected through HPLC [36], GC [37] and enzyme linked immunosorbent assays, using monoclonal and polyclonal antibodies [38]. The application of electrochemical luminescence sensors [39], imprinted polymer sensors [40], carbon modified electrodes [41] and sol-gel immunosorbents [42] has also been examined for low level IPU detection.

Nanocomposites play different roles in nanosensors construction process, with their different compositions, morphologies and sizes. Due to the unique nanocomposites properties, they have showed significant enhancement effects on the EC determination of different analytes [43-45]. Nanocomposites have been incorporated into electrochemical sensors by the WE surface modification, and used for the investigation of various pesticides. In this study, a PMMA/M(FexOy) PNC modified GCE was developed as WE, and employed for IPU quantification, using CV and SWV EC methods. Nano M(FexOy) has been used as a very powerful determination material for biomolecules detection. Herein, PMMA/M(FexOy) PNC has been employed for the first time as a detection material for IPU quantification. Due to the M(FexOy) filler uniform dispersion and high conducting nature in PMMA, PNC/GCE exhibited good CV and SWV responses, and it is proposed as a reasonable alternative for IPU quantification. The IPU detection protocol through this work may also be used for the EC detection of different pesticides in environmental samples.

Materials and methods

Reagents

IPU, with Arelon trade name (75%, Aventis Crop Science India, Ltd.), was crystallized into ethanol, at the melting point (mp) of 157.5 °C. Other chemicals and solvents (with a purity of >98%) were indigenously procured as analytical reagent grades, and used without further purification. All aqueous solutions were prepared with distilled water.

Ferrite Polymer Nanocomposites (FPNCs) were prepared and characterized according to their crystallite size, through X-ray Diffraction (XRD), following earlier reported procedures [46]. Methylmethacrylate (MMA) was purified through the inhibitor extraction into sodium hydroxide (NaOH: 10%, w/v), followed by distillation under reduced pressure, at the boiling point (b.p.) of 98 °C/10 mm/Hg (mercury), d of 0.94 g/cc (cubic centimeter) and λ_{\max} (wavelength of maximum absorbance) of 2.48 nm (nanometer) [47]. PMMA/M(FexOy) PNC was prepared through MMA (2.5×10^{-3} mol/dL) 2,2'-azobisisobutyronitrile (8.25×10^{-3} mol/dL) initiated polymerization, with FPNCs (0.5%, w/v), at 1200 psi (pounds per square inch), 70 ± 1 °C, for 6 h, in supercritical CO₂ (carbon dioxide) [48]. All the electrochemical experiments were conducted at IPU concentrations ranging from 0 to 16.68 ng/dL, in 1 M HClO₄, under ambient temperature. The pH of the medium during experiments was varied from 2.0 to 9.0.

Instrumentations

PNC/GCE SEM images were recorded on JEOL, JSM 6610 LV, at 0.2 KX (7 μ m: micrometer) and 15 kV (kilovolt), under identical conditions, for comparable results.

AFM images were scanned under a tapping mode, over NTEGRA Prima, through ultra sharp Si cantilevers (48 N/m force constant). IPU EC studies were performed over an IVIUM potentiostat-galvanostat, using a triple electrode cell assembly.

WE fabrication

Firstly, a GCE was thoroughly finished with a fine aluminum slurry on a diamond pad, followed by ultrasonicated cleaning with acetone. A homogenous solution of coating material was prepared by heating PMMA/M(FexOy) and GO (40%) in tetrahydrofuran (1 mg/mL), at 50 ± 1 °C, for 20 min, followed by sonication (30 min). Then, the bare GCE top surface (3 mm diameter) was modified through its dip coating by the prepared suspension, followed by its drying at 50 ± 1 °C, 400 mmHg, for 1 h.

Electrochemical measurements

All the EC experiments were conducted at IPU concentrations ranging from 0 to 16.68 ng/dL, in HClO₄ (1 M), at pH 2 [49 - 50], under ambient temperature. GCE and PNC/GCE, a Pt foil (1 cm²) and Ag/AgCl were used as working, auxiliary and reference electrodes, respectively. The developed electrodes EC for IPU detection was conducted through CV, and the peak currents (μA) and peak potentials (V) were measured. CV was scanned under the potential range from 1.55 to -1.15 V of 0.045 V/s. IPU was quantified with SWV, by plotting a calibration graph between the maximum peak current and the respective increasing concentrations. The experimental parameters (pH of the medium, supporting electrolyte (s.e.), SWV frequency, pulse amplitude and step potential) were optimized in relation to the maximum peak current and selectivity. The SWV conditions were optimized at the intervals of 5 mV, accumulation potential of - 0.3 V, frequency of 35 Hz and amplitude of 30 mV. The LOD and LOQ were calculated from the SWV calibration curve using the formula: $LOQ = 10 s/m$ and $LOD = 3.3 s/m$, where *s* is the intercept and *m* is the slope of the calibration curve.

Results and discussion

PNC/GCE surface characterization

The transfer of electrons from the coated conductive polymer matrix to the electrode is one of the key parameters in EC determination which depends on the stability and morphology of the coating material over the bare electrode. In this study, PNC/GCE surface characterization was carried out by SEM and AFM studies. SEM revealed that PMMA had a phase separated morphology, due to the tetrahydrofuran (THF) non-uniform expulsion [51]. PNC displayed a well distinct biphasic morphology corresponding to the homogeneous PMMA phase, with occasional FPNCs clusters (Fig. 1a and 1b) [52, 53].

A further insight into FPNCs qualitative dispersion into the PMMA matrix was made through AFM, which revealed its homogeneous surface texture, with an average roughness of 1.69 nm (Fig. 1c). This was associated with occasional defects, due to the solvent evaporation during casting procedures. FPNCs dispersion into the PMMA matrix afforded PNCs with an average roughness of 7.35 nm, and an approximate particle size ranging from 17 to 400 nm (Fig. 1d). The two phase microstructure, consisting of a light matrix and relatively dark interstitial regions, indicates that PNCs interface side was heterogeneous. This accounts for the increase in PNCs average roughness over

the GCE surface. The PMMA film shows that the average holes depth in PNC was 12.5 nm (Fig 1c). FPNCs reinforcement into the PMMA matrix has increased them to 25 nm (Fig. 1d).

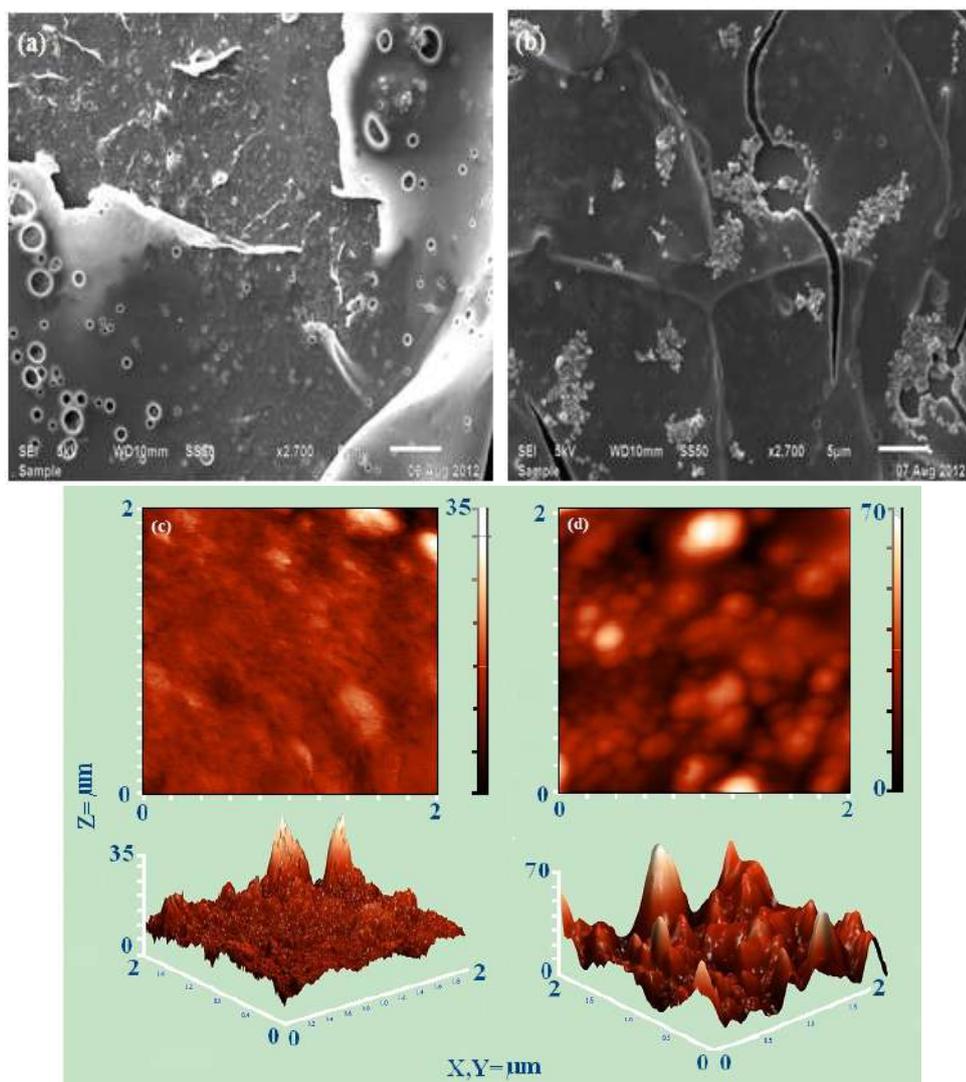


Figure 1. SEM of (a) PMMA; and (b) PNC. AFM of (c) PMMA; and (d) PNC.

IPU electrochemical behavior at PNC/GCE

The GCE is a key factor in the course of IPU EC, and its properties are governed by the nature of the modifying adsorbent material coating surface. In this work, PNC/GCE was found to provide good electron transfer properties and accessible electrode potential ranges, developing a highly stable WE for IPU monitoring. In PNC, the PMMA percentage in PMMA/M(FexOy) had a pronounced influence on IPU adsorption and, consequently, on its voltammetric response. Different PMMA and PMMA/M(FexOy) compositions (20%, 30% and 40% by wt.) were examined, to record the best signal response. Maximum current was attained for 40% by wt. of PMMA in PMMA/M(FexOy) PNC. There was a notable decrease in the peak current for the PMMA higher concentrations. The probable reason for that is the conducting M(FexOy) particle reduction at the modified GCE surface.

An optimized PNC coating was applied over GCE, and the working conditions were maximized to obtain the best results for IPU EC monitoring. A thorough study was carried out with a broad range of possible electrolytes, including HCl (hydrochloric acid), H₂SO₄ (sulfuric acid), HNO₃ (nitric acid) and HClO₄, to evaluate the s.e. Optimum pH and best EC results for IPU were obtained with 1 M HClO₄, at pH 2, at both GCE and PNC/GCE.

IPU CV studies at PNC/GCE

CV studies were performed to verify IPU electro-activity at the WE, in 1 M HClO₄, at pH 2, under ambient temperature. Fig. 2 depicts IPU CV (0.5 μM), in a potential range from 0.15 to 1.5 V, at a scan rate of 25 mV/s. The CV shape indicates a quasi-reversible redox response for the adsorbed IPU at GCE and PNC/GCE, in the used conditions. The CV shows cathodic peaks at 1.08 V, which relate to electro-reduction, and an anodic peak at 1.35 V, which corresponds to IPU electro-oxidation. It was found that the magnitude of cathodic and anodic peak currents produced for IPU at PNC/GCE was much higher than that of the bare GCE, due to the PNC good electrical conductivity and high surface area, which are believed to enhance electrons transfer. IPU CV at PNC/GCE reflected sharp redox peaks with a cathodic peak current (I_{pc}) and an anodic peak current (I_{pa}) at about -34 μA and 80 μA, respectively (Fig. 2).

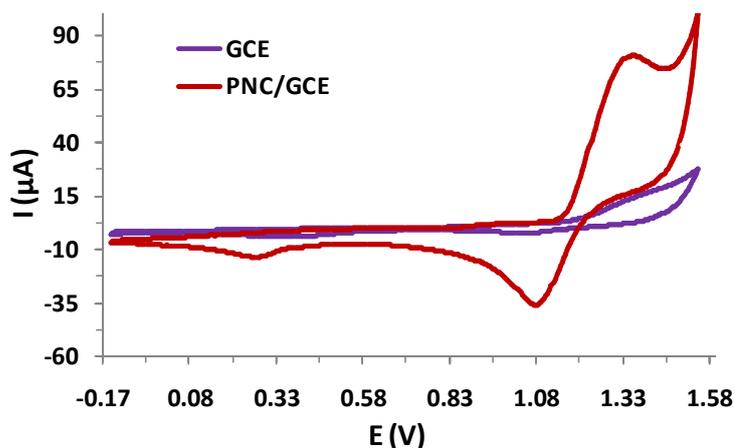


Figure 2. IPU CV on GCE and PNC/GCE in 1 M HClO₄.

Scan rate effect

Fig. 3 shows the scan rate effect on the IPU CV response at PNC/GCE in 1 M HClO₄, with a gradual increase of 15, 25, 50, 75, 100, 200, 300 and 400 mV/s, respectively. The redox peak currents showed a good linear relationship with the scan rate, in the potential range from 0.2 to 1.6 V.

It was seen that the peak potential did not vary with the scan rate, and that the oxidation peak height greatly increased with higher scan rates, from 15 mV/s to 400 mV/s. However, the reduction adsorption peak height did not significantly vary with an increase in the scan rate. This may be due to the ohmic drop and greater capacitive currents. Since it was found that a scan rate of 200 mV/s was best suited for IPU scanning in other optimized parameters, it was selected for all further studies.

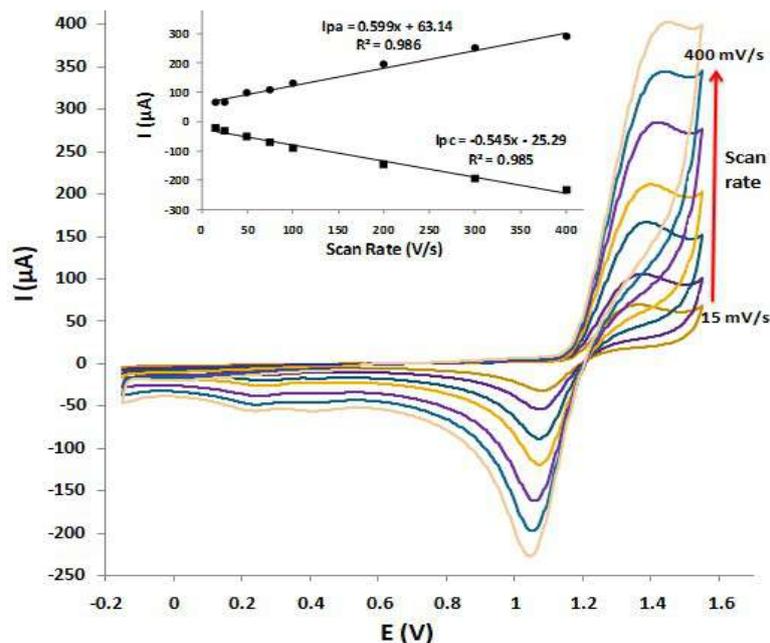


Figure 3. IPU CVs of on PNC/GCE, at a scan rate varying from 15 mV to 400 mV, in 1 M HClO₄ (pH 2).

IPU concentration effect on CV

The higher IPU concentrations effect on CV was investigated in 1 M HClO₄ (pH 2), at the developed PNC/GCE. As shown in Fig. 4, a uniform CV pattern was attained with successive increments in IPU concentrations, from 0.0 to 1.2 μM . Each subsequent IPU addition up to 1.2 μM reflected an increase in the peak current, which indicated a sensitive IPU detection by the PNC/GCE.

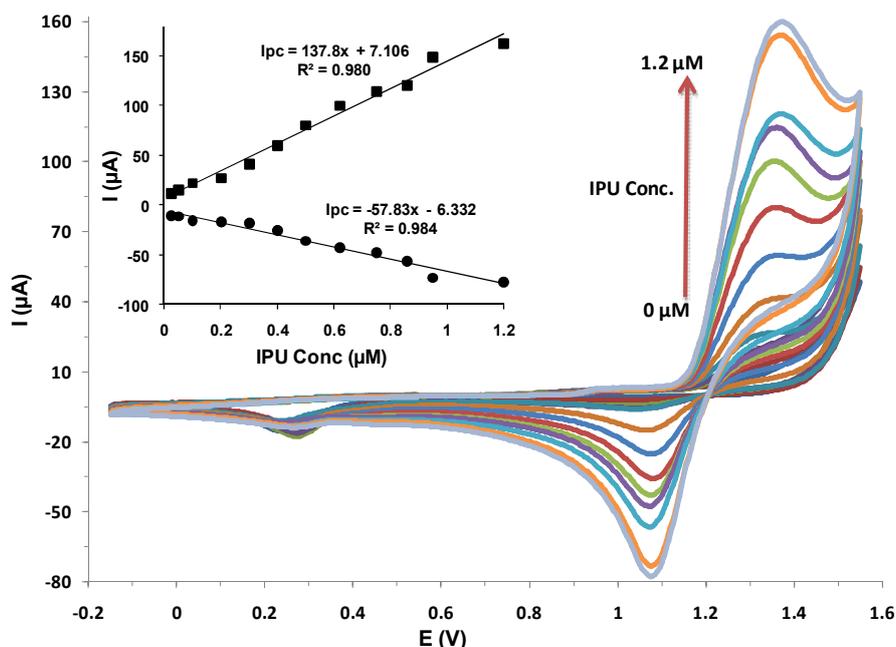


Figure 4. IPU CV on PNC/GCE in 1 M HClO₄ (pH 2), with successive increases in concentration from 0.01, 0.025, 0.05, 0.075, 0.10, 0.20, 0.30, 0.40, 0.60, 0.80, 1.0 to 1.20 μM , respectively.

The CV indicated that, at higher sides, the peaks were sharp, but a response plateau appeared after IPU 1.2 μM concentration, while, at the lower end, the peaks were not significant at an IPU concentration below 0.025 μM . On the basis of these obtained results, a linear relationship was established between the peak currents and the increase in IPU concentrations.

IPU SWV at PNC/GCE

In order to check the viability of the developed PNC/GCE for IPU estimation, electrochemical parameters were optimized in SWV. The best response was obtained in the potential range from 0 to 3.0 V, with an accumulation time of 50 s, a SW frequency of 35 Hz, a SW step potential of 15 mV and an SW amplitude of 30 mV, respectively. On the successive addition of IPU into a 1 M HClO_4 (pH 2) solution, a uniform increase in the peak current was found with successive higher IPU concentrations. The SWV peak currents were recorded using optimized parameters for a series of IPU concentrations ranging from 0 to 1.2 μM (Fig. 5).

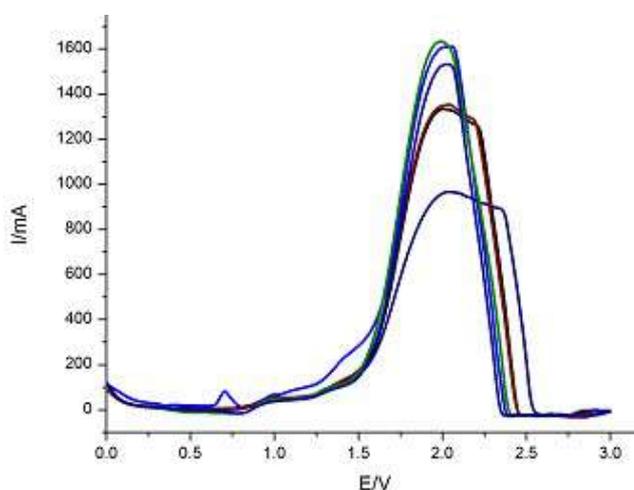


Figure 5. IPU SWV on PNC/GCE with concentrations ranging from 0.025 to 1.2 μM , in 1 M HClO_4 , at pH 2.

A well-defined linear calibration curve was obtained between the peak current and IPU concentration at PNC/GCE, with a LOQ of 1.98×10^{-7} M and a LOD of 6.5×10^{-8} M, respectively (Fig. 6). The SWV results specify that PNC/GCE has an excellent reproducibility for IPU estimation, which indicates that PMMA/M(FexOy) modified electrodes can be employed for electrochemical detection purposes, as an alternative source of analysis to expensive detection techniques. This makes the proposed SWV method based on PNC/GCE a fast, sensitive and better standard method. LOD and LOQ values attained by this method are significantly better than those of earlier reported methods, which indicates that SWV could be employed to analyze IPU in different environmental samples.

Interference study

Interference studies were performed to verify the applicability of PNC/GCE proposed methodology for IPU selective estimation. The existence of certain

important ions in natural samples could be a probable source of interference in IPU analysis by SWV. Thus, the influence of a range of sodium, hydrochloric acid, sulfate, nitrate, magnesium, calcium, iron, zinc, potassium and copper ions (Na^+ , Cl^- , SO_4^{2-} , NO_3^- , Mg^{2+} , Ca^{2+} , Fe^{3+} , Zn^{2+} , K^+ and Cu^{2+} , respectively) on the PNC/GCE electrochemical sensor was examined by the introduction of the ions salts in the IPU solution. The PNC/GCE tolerance limit was evaluated by increasing the interfering ions concentrations at a constant IPU concentration ($5 \mu\text{M}$). A comparison of amperometric responses showed a higher peak current value for IPU than that of other interfering ions (Fig. 7).

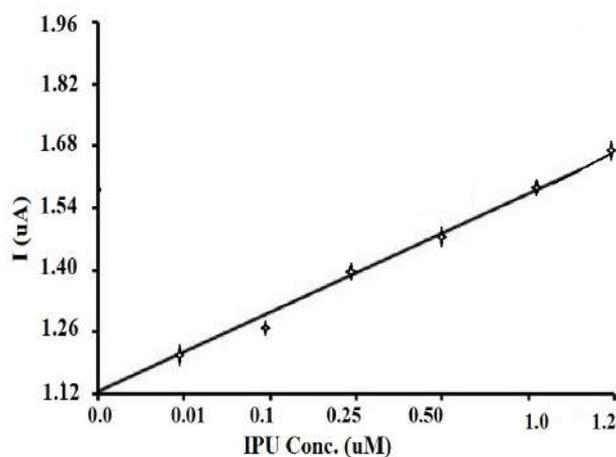


Figure 6. SWV calibration plot with successive increases in IPU concentrations.

The outcomes revealed no significant interfering effect of the respective ions on IPU determination up to $1.4 \mu\text{M}$. This confirms the SWV better selectivity for simultaneous IPU detection in a wide range of environmental samples.

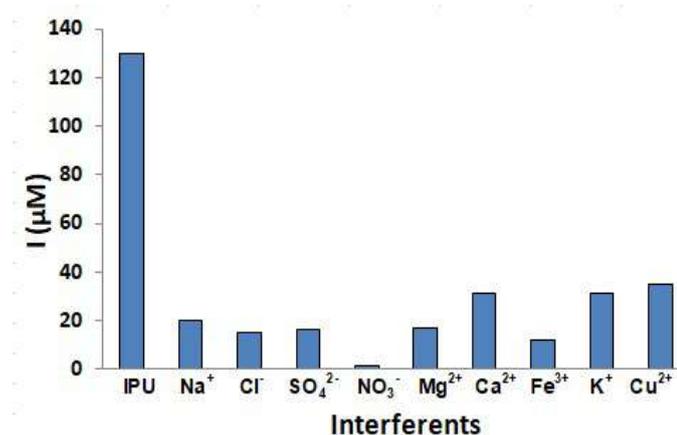


Figure 7. Comparative amperometric response of common interfering ions on IPU estimation at PNC/GCE.

Conclusions

Electrochemical techniques are currently the leading and most promising probes for pesticides monitoring. The prime goal of the present study was to establish a reasonable and sensitive means of IPU analysis. PMMA/M(Fe_xO_y) modified GCE (PNC/GCE), as WE, was developed, characterized and employed for IPU

electrochemical detection. CV and SWV procedures for PNC/GCE provide convenient and efficient methods for IPU electrochemical estimation. CVs indicate that the current response obtained on PNC/GCE was quite better than that of the bare GCE. Therefore, the PMMA/M(FexOy) modified WE was selected for IPU monitoring. SWV studies showed IPU sensitive and selective estimation, with a LOQ of 1.98×10^{-7} M and a LOD of 6.5×10^{-8} M, without any significant interference of common coexisting ions. Additionally, a careful analysis of the results obtained from quantification limits reveals that PNC/GCE delivers an alternative method for IPU electrochemical detection. The developed protocol suggested a possibility of its application to the analytical determination of other pesticides that are usually detected at mercury electrodes with the disadvantages of being costly and more time consuming.

Acknowledgements

The authors acknowledge the Department of Chemistry, G. B. Pant University of Agriculture and Technology, Pantnagar, India, for providing all the support during the study period.

Authors' contribution

Sameena Mehtab: wrote the paper; performed data validation. **M. G. H. Zaidi:** conceived and designed the analysis; performed conceptualization and supervision. **Priya Bhatt:** performed the analysis; performed laboratory work and formal analysis. **Pragati Joshi:** collected the data; draw the manuscript images. **Tithi Agarwal:** contributed with data or analysis tools: work station and software handling.

References

1. El Mhammedi MA, Bakasse M, Bachirat R, et al. Square wave voltammetry for analytical determination of paraquat at carbon paste electrode modified with fluoroapatite. *Food Chem.* 2008;110(4):1001-1006. DOI: <https://doi.org/10.1021/jf9800357>
2. MacDonald LM, Meyer TR. Determination of imidacloprid and triadimefon in white pine by gas chromatography/mass spectrometry. *J Agri Food Chem.* 1998;46(8):3133-3138. DOI: <https://doi.org/10.1021/jf9800357>
3. Valenzuela AI, Lorenzini R, Redondo MJ, et al. Matrix solid-phase dispersion microextraction and determination by high-performance liquid chromatography with UV detection of pesticide residues in citrus fruit. *J Chrom A.* 1999;839(1-2):101-107. DOI: [https://doi.org/10.1016/S0021-9673\(99\)00055-2](https://doi.org/10.1016/S0021-9673(99)00055-2)
4. Lehotay SJ, Kok AD, Hiemstra M, et al. Validation of a fast and easy method for the determination of residues from 229 pesticides in fruits and vegetables using gas and liquid chromatography and mass spectrometric detection. *J AOAC International.* 2005;88(2):595-614. DOI: <https://doi.org/10.1093/jaoac/88.2.595>
5. Aguilar C, Borrull F, Marce RM. Determination of pesticides in environmental waters by solid-phase extraction and gas chromatography with electron-capture

- and mass spectrometry detection. *J Chrom A*. 1997;771(1-2):221-231. DOI: [https://doi.org/10.1016/S0021-9673\(97\)00091-5](https://doi.org/10.1016/S0021-9673(97)00091-5)
6. Viswanathan S, Radecka H, Radecki J. Electrochemical biosensor for pesticides based on acetylcholinesterase immobilized on polyaniline deposited on vertically assembled carbon nanotubes wrapped with DNA. *Biosens Bioelectron*. 2009;24(9):2772-2777. DOI: <https://doi.org/10.1016/j.bios.2009.01.044>
 7. Liu K, Dong H, Deng Y. Recent advances on rapid detection of pesticides based on enzyme biosensor of nanomaterials. *J Nanosci Nanotechnol*. 2016;16(7):6648-6656. DOI: <https://doi.org/10.1166/jnn.2016.11392>
 8. Manisankar P, Viswanathan S, Pusphalatha AM, et al. Electrochemical studies and square wave stripping voltammetry of five common pesticides on poly 3, 4-ethylenedioxythiophene modified wall-jet electrode. *Analyt Chem Acta*. 2005;528(2):157-163. DOI: <https://doi.org/10.1016/j.aca.2004.08.027>
 9. Wang H, Casalongue HS, Liang Y, et al. Ni (OH)₂ nanoplates grown on graphene as advanced electrochemical pseudocapacitor materials. *J Am Chem Soc*. 2010;132(21):7472-7477. DOI: <https://doi.org/10.1021/ja102267j>
 10. Brahim MB, Ammar HB, Abdelhédi R, et al. Electrochemical behavior and analytical detection of Imidacloprid insecticide on a BDD electrode using square-wave voltammetric method. *Chinese Chem Lett*. 2016;27(5):666-672. DOI: <https://doi.org/10.1016/j.cclet.2015.12.032>
 11. Reddy Prasad P, Ofamaja AE, Reddy CN, et al. Square Wave Voltammetric Detection of Dimethylvinphos and Naftalofos in Food and Environmental Samples Using RGO/CS modified Glassy Carbon Electrode. *Int J Electrochem Sci*. 2016;11(1):65-79.
 12. Mirceski V, Gulaboski R. Recent achievements in square-wave voltammetry (a review). *Maced J Chem Eng*. 2014;33(1):1-12. DOI: <https://doi.org/10.20450/mjce.2014.515>
 13. Liu G, Lin Y. Electrochemical sensor for organophosphate pesticides and nerve agents using *zirconia* nanoparticles as selective sorbents. *Analyt Chem*. 2005;77(18):5894-5901. DOI: <https://doi.org/10.1021/ac050791t>
 14. Jeyapragasam T, Saraswathi R. Electrochemical biosensing of carbofuran based on acetylcholinesterase immobilized onto iron oxide–chitosan nanocomposite. *Sens Actuat B Chem*. 2014;191:681-687. DOI: <https://doi.org/10.1016/j.snb.2013.10.054>
 15. Zaidi SA, Shin JH. A novel and highly sensitive electrochemical monitoring platform for 4-nitrophenol on MnO₂ nanoparticles modified graphene surface. *RSC Adv*. 2015;5(108):88996-89002. DOI: <https://doi.org/10.1039/c5ra14471j>
 16. Manisankar P, Selvanathan G, Vedhi C. Utilization of sodium montmorillonite clay-modified electrode for the determination of isoproturon and carbendazim in soil and water samples. *Appl Clay Sci*. 2005;29(3-4):249-257. DOI: <https://doi.org/10.1016/j.clay.2005.01.006>
 17. Noyrod P, Chailapakul O, Wonsawat W, et al. The simultaneous determination of isoproturon and carbendazim pesticides by single drop analysis using a graphene-based electrochemical sensor. *J Electroanal Chem*. 2014;719:54-59. DOI: <https://doi.org/10.1016/j.jelechem.2014.02.001>

18. Wu B, Hou L, Du M, et al. A molecularly imprinted electrochemical enzymeless sensor based on functionalized gold nanoparticle decorated carbon nanotubes for methyl-parathion detection. *RSC Adv.* 2014;4(96):53701-53710. DOI: <https://doi.org/10.1039/c4ra06210h>
19. Manisankar P, Sundari PA, Sasikumar R, et al. Electroanalysis of some common pesticides using conducting polymer/multiwalled carbon nanotubes modified glassy carbon electrode. *Talanta.* 2008;6(5):1022-1028. DOI: <https://doi.org/10.1016/j.talanta.2008.04.056>
20. Long X, Deng C, Xiao G, et al. Electrochemical Sensors with Antifouling Properties for Sensitive Detection of Isoproturon Based on Glassy Carbon Electrode Modified with Nafion Membrane. *Int J Electrochem Sci.* 2020;15:4964-4977. DOI: <https://doi.org/10.20964/2020.06.58>
21. Wang M, Huang J, Wang M, et al. Electrochemical nonenzymatic sensor based on CoO decorated reduced graphene oxide for the simultaneous determination of carbofuran and carbaryl in fruits and vegetables. *Food Chem.* 2014;151:191-197. DOI: <https://doi.org/10.1016/j.foodchem.2013.11.046>
22. Zhou Q, Yang L, Wang G, et al. Acetylcholinesterase biosensor based on SnO₂ nanoparticles-carboxylic graphene-nafion modified electrode for detection of pesticides. *Biosens Bioelectron.* 2013;49:25-31. DOI: <https://doi.org/10.1016/j.bios.2013.04.037>
23. Li Y, Xu M, Li P, et al. Nonenzymatic sensing of methyl parathion based on graphene/gadolinium Prussian Blue analogue nanocomposite modified glassy carbon electrode. *Analyt Met.* 2014;6(7):2157-2162. DOI: <https://doi.org/10.1039/c3ay41820k>
24. Ebrahim S, El-Raey R, Hefnawy A, et al. Electrochemical sensor based on polyaniline nanofibers/single wall carbon nanotubes composite for detection of malathion. *Synth Met.* 2014;190:13-19. DOI: <https://doi.org/10.1016/j.synthmet.2014.01.021>
25. C. Tomlin (Ed). *The Pesticide Manual*. Royal Society of Chemistry and the British Crop Protection Council. Farnham. Surrey. UK. 1994.
26. Kramer PM, Goodrow MH, Kremmer E. Enzyme-linked immunosorbent assays based on rabbit polyclonal and rat monoclonal antibodies against isoproturon. *J Agri Food Chem.* 2004;52(9):2462-2471. DOI: <https://doi.org/10.1021/jf035498d>
27. European commission. 2000 European commission EU water framework directive 2000/60/EC.
28. Varshney S, Hayat S, Alyemeni MN, et al. Effects of herbicide applications in wheat fields: is phytohormones application a remedy? *Plant Sign Beh.* 2012;7(5):570-575. DOI: <https://doi.org/10.4161/psb.19689>
29. Yin XL, Jiang L, Song NH, et al. Toxic reactivity of wheat (*Triticum aestivum*) plants to herbicide isoproturon. *J Agri Food Chem.* 2008;56(12):4825-4831. DOI: <https://doi.org/10.1021/jf800795v>
30. Sarkar SN, Majumdar AC, Chattopadhyay SK. Effect of isoproturon on male reproductive system: clinical, histological and histoenzonological studies in rats. *Ind J Exp Biol.* 1997;35(2):133-138.

31. Peres F, Florin D, Grollier T, et al. Effects of the phenylurea herbicide isoproturon on periphytic diatom communities in freshwater indoor microcosms. *Environ Poll.* 1996;94(2):141-152. DOI: [https://doi.org/10.1016/S0269-7491\(96\)00080-2](https://doi.org/10.1016/S0269-7491(96)00080-2)
32. Chauhan LK, Kumar M, Paul BN, et al. Cytogenetic effects of commercial formulations of deltamethrin and/or isoproturon on human peripheral lymphocytes and mouse bone marrow cells. *Environ Mol Mutagen.* 2007;48(8):636-643. DOI: <https://doi.org/10.1002/em.20330>
33. Zaidi MGH, Agarwal T, Alam S, et al. Synthesis of Fullerene Polyvinyl Pyridine Composites in Supercritical Carbon Dioxide. *Fuller Nanotub Carb N.* 2011;19(4):329-342. DOI: <https://doi.org/10.1080/15363831003721799>
34. Mou RX, Chen MX, Zhi JL. Simultaneous determination of 15 phenylurea herbicides in rice and corn using HPLC with fluorescence detection combined with UV decomposition and post-column derivatization. *J Chrom B.* 2008;875(2):437-443. DOI: <https://doi.org/10.1016/j.jchromb.2008.09.022>
35. Singh KP, Prajapati RK, Ahlawat S, et al. Use of isoproturon imprinted polymer membranes as a selective recognition platform in a resistance based electrochemical sensor. *Open J Appl Biosens.* 2013;2(1):28317. DOI: <https://doi.org/10.4236/ojab.2013.21003>
36. Ruberu SR, Draper WM, Perera SK. Multiresidue HPLC methods for phenyl urea herbicides in water. *J Agri Food Chem.* 2000;48(9):4109-4115. DOI: <https://doi.org/10.1021/jf000266p>
37. Ferrer I, Pichon V, Hennion MC, et al. Automated sample preparation with extraction columns by means of anti-isoproturon immunosorbents for the determination of *phenylurea* herbicides in water followed by liquid chromatography–diode array detection and liquid chromatography–atmospheric pressure chemical ionization mass spectrometry. *J Chrom A.* 1997;777(1):91-98. DOI: [https://doi.org/10.1016/S0021-9673\(97\)00437-8](https://doi.org/10.1016/S0021-9673(97)00437-8)
38. Gerecke AC, Tixier C, Bartels T, et al. Determination of *phenylurea* herbicides in natural waters at concentrations below 1 ng l⁻¹ using solid-phase extraction, derivatization, and solid-phase microextraction–gas chromatography–mass spectrometry. *J Chrom A.* 2001;930(1-2):9-19. DOI: [https://doi.org/10.1016/S0021-9673\(01\)01192-X](https://doi.org/10.1016/S0021-9673(01)01192-X)
39. Li J, Li Y, Zhang Y, et al. Highly sensitive molecularly imprinted electrochemical sensor based on the double amplification by an inorganic prussian blue catalytic polymer and the enzymatic effect of glucose oxidase. *Analyt Chem.* 2012;84(4):1888-1893. DOI: <https://doi.org/10.1021/ac2026817>
40. Sadriu I, Bouden S, Nicolle J, et al. Molecularly imprinted polymer modified glassy carbon electrodes for the electrochemical analysis of isoproturon in water. *Talanta.* 2020;207:120222. DOI: <https://doi.org/10.1016/j.talanta.2019.120222>
41. Amra S, Bataille T, Bourouina Bacha S, et al. Nanostructured Modified Carbon Paste Electrode as Voltammetric Sensor for Isoproturon Trace Analysis in Water. *Electroanalysis.* 2020;32(6):1346-1353. DOI: <https://doi.org/10.1002/elan.201900083>

42. Zhang X, Martens D, Kramer PM, et al. Development and application of a sol–gel immunosorbent-based method for the determination of isoproturon in surface water. *J Chrom A*. 2006;1102(1-2):84-90. DOI: <https://doi.org/10.1016/j.chroma.2005.10.072>
43. Joshi P, Mehtab S, Tyagi T, et al. Development of polyindole/tungsten carbide nanocomposite-modified electrodes for electrochemical quantification of *chlorpyrifos*. *J Nanostr Chem*. 2019;10:33-45. DOI: <https://doi.org/10.1007/s40097-019-00326-9>
44. Sharma S, Joshi P, Mehtab S, et al. Development of non-enzymatic cholesterol electrochemical sensor based on polyindole/tungsten carbide nanocomposite, *J Analyt Test*. 2020;4:13-22. Doi: <https://doi.org/10.1007/s41664-020-00117-0>
45. Rita, Mehtab S, Zaidi MGH, et al. Polyindole Based Nanocomposites and their Applications: A Review. *Mat Sci Res India*. 2019;16 (2):97-102. DOI: <http://dx.doi.org/10.13005/msri/160202>
46. Zaidi MGH, Sah PL, Alam S, et al. Synthesis of epoxy ferrite nanocomposites in supercritical carbon dioxide *J Exp Nanosci*. 2009;4(1):55-66. DOI: <https://doi.org/10.1080/17458080802656515>
47. Joshi SK, Kapil JC, Rai AK, et al. Quantitative examination of polymethyl methacrylate graft polybisphenol-A-carbonate copolymer by photoacoustic spectroscopy. *Physi Stat Solidi (a)*. 2003;199(2):321-328. DOI: <https://doi.org/10.1002/pssa.200306640>
48. Bisht G, Zaidi MGH. Supercritical synthesis of poly (2-dimethylaminoethyl methacrylate)/ferrite nanocomposites for real-time monitoring of protein release. *Drug Del Translat Res*. 2015;5(3):268-274. DOI: <https://doi.org/10.1007/s13346-015-0225-3>
49. Nasir T, Gamero-Quijano A, Despas C, et al. Signal amplification by electro-oligomerisation for improved isoproturon detection. *Talanta*. 2020;220:121347. DOI: <https://doi.org/10.1016/j.talanta.2020.121347>
50. Zhou Y, Cheng F, Hong Y, et al. Rapid and Sensitive Detection of Isoproturon Via an Electrochemical Sensor Based on Highly Water-Dispersed Carbon Hybrid Material. *Food Analyt Methods*. 2020;13:1-11. DOI: <https://doi.org/10.1007/s12161-020-01707-5>
51. Cao Z, Jiang W, Ye X, et al. Preparation of superparamagnetic Fe₃O₄/PMMA nano composites and their magnetorheological characteristics. *J Mag Magnetic Mat*. 2008;320(8):1499-1502. DOI: <https://doi.org/10.1016/j.jmmm.2007.12.007>
52. Roy D, Kumar PS. Enhancement of (BH) max in a hard-soft-ferrite nanocomposite using exchange spring mechanism. *J Appl Phys*. 2009;106(7):3902-3909. DOI: <https://doi.org/10.1063/1.3213341>
53. Vollath D, Szabó DV. Synthesis and properties of nanocomposites. *Adv Eng Mat*. 2004;6(3):117-127. DOI: <https://doi.org/10.1002/adem.200300568>