Electrochemical Behavior Study of Sodium Saccharin in an Aqueous Electrolyte Solution by a Nano-sensor, Using Cyclic Voltammetric Technique

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Abstract

This study investigated one of the chemical compounds taken by diabetic patients as a replacement for natural sugar. Its aim was to identify the electrochemical properties of sodium saccharin in different electrolytes, using a nano-sensor. Sodium saccharine was studied by cyclic voltammetric technique, in different electrolytes, using a modified glassy carbon electrode (GCE) with carbon nanotubes (CNT) as working electrode (CNT/GCE). It was found that the redox current peaks of 0.01 mM sodium saccharine in 1 M Na₂SO₃ enhanced both redox peaks with the CNT electro-catalyst on the GCE surface. Different concentrations, pH and scan rates of sodium saccharine in Na₂SO₃ have been studied. Also, the nano sensor showed good reliability and stability towards the chemical compounds in the cyclic voltammetric cell. Other electrochemical parameters were determined, such as the potential peak separation (Epa-Epc \approx 100 mV), the current ratio (Ipa/Ipc \approx 1) of the redox peaks and the cathodic-anodic reaction rate. The diffusion coefficient value was determined at different scan rates.

Keywords: sodium saccharine, cyclic voltammetry, CNT/GCE and Na₂SO₃.

Introduction

Many different studies have covered the chemical effects of redox current peaks on several problems in the human body by oxidative action, using electrochemical methods¹⁻⁵.

Sodium saccharin is a non-nutritive sweetener, which means that it is not metabolized by the body, to produce energy⁶. The redox current peaks of complex systems of manganese in saccharine Mn–saccharine and Mn–saccharine–phenanthroline were studied by cyclic voltammetry, using GCE in 0.1 M KCl. It was found that the Mn-Saccharine complex is irreversible and quasi-reversible of the Mn–saccharine–phenanthroline⁷ complex. Saccharin was studied on an Au electrode in 0.1 M HClO₄ as electrolyte, by cyclic voltammetric

technique. It was found that the stability of the adsorbed monolayer was enhanced by an increase in the saccharin concentration, and by a decrease in temperature⁸. Saccharin was studied with Fe(III) as a complex reagent [Fe(C₇H₄O₃SN)₂Cl]. The infrared and electronic spectral spectroscopy have found that two saccharin molecules were linked to the Fe(III) ion as a complex. Also, a quasi-reversible electron transfer process of the iron complex was studied by cyclic voltammetric technique⁹. Four sweeteners – sodium saccharin, aspartame, sodium cyclamate and acesulfame-K – were studied, and the average recoveries for various samples ranged from 93 to 107%¹⁰. The cyclic voltammograms of the chemical Schiff-base produced two cathodic current peaks in Britton-Robinson buffer (pH 7.0-10.0). The potential peaks values ranged from 0.0 V to -1.4 V. The cathodic peaks at more positive potentials than the reduction peaks of saccharin may be assigned to the cathodic reaction of C-N+ and >C=N- moieties of Schiff-base¹¹. The oxidation-reduction current peaks properties of the Fe(III)/Fe(II) reaction were found to be reversible, using a platinum electrode with the-system ligands, and also, quasi-reversible in the reaction¹². Saccharin was used in an electrolyte solution, by an electrochemical quartz crystal microbalance (EQCM) technique, coupled with cyclic voltammetry (CV) measurements. The oxidation-reduction reactions from potentiodynamic polarization experiments were in agreement with the EQCM results¹³.

In this work, sodium saccharin was studied, as a sugar substitute, by the cyclic voltammetric method, to determine its the electrochemical behavior in different chemical properties at different electrolyte solutions, and a nano-sensor was used as electro-catalyst working electrode.

Experiment

Materials

Sodium saccharin (purity 98%), from China, carbon nanotubes (purity 99%) supplied from Fluka company (Germany), and other chemicals used in the experiment, such as KCl, K₂HPO₄, KH₂PO₄, Na₂SO₃, KClO₄, K₂SO₄ and KNO₃ (in high purity materials), from SCRC (China) were used. Deionized water was used for the preparation of aqueous solutions. Buffer solutions had sodium acetate with acetic acid, for an acidic pH, and sodium hydroxide, for an alkaline pH. All solutions used in the cyclic voltammetric cell were treated with nitrogen gas for 10-15 minutes, prior to being oxygen-free.

Apparatus

An EZstat series (Potentiostat/Glvanostat) NuVant Systems Inc (made in USA) instrument was used in all experiments. The electrochemical bio-analytical cell was connected to a potentiostat device, and monitored through the special program that has been installed on the personal computer to perform cyclic voltammetry (CV). A silver-silver chloride electrode (Ag/AgCl in 3M NaCl) and a platinum wire (1 mm diameter) were as reference and counter electrodes, respectively. A glassy carbon working electrode (GCE) modified with CNT was used in this study, after being cleaned with an aluminum solution, and treated

with ultrasonic path water for ten minutes. All experiments were done at the room temperature of 25 $^{\circ}$ C.

Preparing the modification of GCE with CNT (CNT/GCE)

A mechanical attachment technical method to prepare the CNT/GCE working electrode was employed to obtain the nano-sensor^{14,15}. The GCE modification method included: the abrasive application of multiwall carbon nanotubes (MWCNT) on the clean GCE surface, forming the MWCNT/GCE modified working electrode; putting it in 10 mL of the electrolyte in the cyclic voltammetric cell; then, connecting it to all electrodes with the potentiostat.

Results and discussion

Effect of different supporting electrolytes

The effect of different supporting electrolytes – KCl, normal saline NaCl 0.9% (NS), K₂HPO₄, KH₂PO₄, Na₂SO₃, KClO₄, K₂SO₄ and KNO₃ – was studied on the oxidation and reduction current peaks of sodium saccharin, using a scan rate of 100 mV/sec, with Ag/AgCl as reference electrode. Table 1 shows the redox current peak of sodium saccharin, at CNT/GCE, as a modified working electrode in different electrolyte solutions. Table 2 shows oxidation peak and redaction peak values for sodium saccharin on the GCE, in different electrolyte solutions. Na₂SO₃ was the best electrolyte solution which enhanced the redox current peaks of sodium saccharin, and it was used in all the experiments, as shown in Fig. 1.



Figure 1. Cyclic voltammogram of 1 mM sodium saccharin at CNT/GCE in 0.1 M Na₂SO₃ and 0.1 M K₂SO₄ electrolyte solutions with pH=3, at 100 mV sec⁻¹, versus Ag/AgCl.

Tables 1 and 2 show the enhancement of redox current peaks in the following orders:

Ipc $_{CNT/GCE}/Ipc _{GCE}$: Na₂SO₃ >KCL>K₂SO₄>K₂HPO₄ >KH₂PO₄ > KCLO₄ ≥NS >KNO₃;

Ipa_{CNT/GCE}/Ipa_{GCE}: Na₂SO₃>KCL>KNO₃ > KH₂PO₄ > NS>K₂HPO₄>KCLO₄> K₂SO₄.

Electrolyte	Ipa, μA CNT/GCE	Ipa, µA GCE	Epa, mV CNT/GCE	Epa, mV GCE	Ipacnt/gce/Ipagce enhancement
KCL	47.29	18.5	59.09	367	2.556
KCLO ₄	18.5	13.8	250	338	1.34
K_2SO_4	16.2	18.2	285	344	0.89
Na ₂ SO ₃	582.5	184	102.9	67.7	3.165
KH ₂ PO ₄	19.5	13.3	333	381	1.466
K ₂ HPO ₄	21.2	19.9	283	270	1.065
KNO ₃	55.8	33.1	912	875	1.685
NS	15.7	14.6	186	516	1.075

Table 1. Current, potential and enhancement values of the oxidation peak current of 1 mM sodium saccharin in different electrolyte solutions, at GCE and CNT/GCE.

Table 2. Current, potential and enhancement values of the reduction current peak of 1 mM sodium saccharin in different electrolyte solutions, at GCE and CNT/GCE.

Electrolyte	Ipc, μA CNT/GCE	Ipc, μA GCE	Epc, mV GCE	Epc, mV CNT/GCE	IPc _{CNT/GCE} /IPc _{GCE} Enhancement
KCL	36.05	17.2	-692	-635	2.556
KCLO ₄	16.9	18.5	-727	-727	1.340
K ₂ SO ₄	20.3	18.7	-668	-645	0.890
Na ₂ SO ₃	547.3	253	-883	-563	3.165
KH ₂ PO ₄	17.9	17.5	-663	-583	1.466
K ₂ HPO ₄	16.9	16.2	-589	-603	1.065
KNO ₃	16.9	30	-648	-551	1.685
NS	16.9	18.5	-710	-658	1.075

Effect of varying pH

The effect of both acidic and alkaline media in Na₂SO₃ for sodium saccharin was studied using a modified GCE with CNT (CNT/GCE) as working electrode, and Ag/AgCl as reference electrode. It was observed that the oxidation-reduction current peaks of sodium saccharin in acidic media gradually linearly increased with lowering pH values, from 6 to 3, as shown in Table 3. Figs. 2 and 3 reveal high current values at pH 3, while both oxidation – reduction current peaks at alkaline pH values, from 7 to 11, decreased to lower values, as shown in Fig. 4. Table 3 shows that the current's Ipa/I_{Pc} ratio value means that the redox reaction of sodium saccharin is a reversible process. Also, the other evidence for the reversibility of the redox reaction of sodium saccharin is the peak separation value which is nearly equal to 100 mV, as shown in Table 3¹⁶. The mechanism of the redox process of sodium saccharin was discussed in the following equations¹²:

Cathodic:
$$C_7H_4O_3SN^- + Na^+ + e^- \rightarrow C_7H_4O_3SN$$
 Na (1)

Anodic: $C_7H_4O_3SN^- + Na^+ - e^- \rightarrow C_7H_4O_3SN Na$ (2)

рН	Ipa u A	Epa mV	Ірс и А	Epc mV	Ipa/I _P c	Epa-Epc mV
3	1009	203.4	590	989.1	1.710169	785.7
4	549	160.5	572.3	272.6	0.959287	112.1
5	442	185.1	440	1162	1.004545	976.9
6	108	99.8	192	1260	0.5625	1160.2
7	42.6	37.81	61.9	1343	0.688207	1305.19
7.5	37.1	15.1	41.3	761	0.898305	745.9
8	53.6	57	24.23	958	2.212134	901
8.5	43.4	51.6	17.9	951	2.424581	899.4
9	61.7	78.9	16.5	846	3.739394	767.1
9.5	22.7	63.6	15.7	952	1.44586	888.4
10.5	32.7	111	15.4	951	2.123377	840
11	13.6	120	13.4	952	1.014925	832

Table 3. Current, potential and peak potential separation values of oxidation-reduction peaks of 1 mM sodium saccharin, with different pHs, at CNT/GCE.



Figure 2. Plot oxidation current against the pH (3-11) of 0.01 mM sodium saccharin in 1 M Na₂SO₃, at CNT/GCE.



Figure 3. Plot reduction current against the pH (3-11) of 0.01 mM sodium saccharin in 1 M Na₂SO₃, at CNT/GCE.

Effect of different concentrations

Figs. 5 and 6 show the calibration curves of different concentrations (0.001-0.01 mM) sodium saccharin in 1 M Na₂SO₃. The detection limit for the low concentrations of sodium saccharin analysis at the CNT/GCE was 10^{-3} mM, with an oxidation current sensitivity close to $308.52 \,\mu$ A/mM.



Figure 4. Cyclic voltammogram of 1 mM sodium saccharin at different pHs (3 and 11) on CNT/GCE, at a scan rate of 100 mV sec⁻¹, with Ag/AgCl as reference electrode.



Figure 5. Plot of oxidation current against different concentrations of sodium saccharin in 1 M Na₂SO₃, at CNT/GCE.



Figure 6. Plot of reduction current against different concentrations of sodium saccharin in 1 M Na₂SO₃, at CNT/GCE.

A curvature at a concentration greater than 10^{-3} mM and at a reduction current sensitivity close to 470.07 μ A/mM was observed. The calibration plots were

performed at the CNT/GCE in the sodium saccharin, with a good linearity of the cathodic current, as described by the equation: y = -3739.4X + 470.07, $R^2 = 0.927$.

Table 4 shows concentration values of sodium saccharin cathodic and anodic current peaks.

Concentration	Ipa	Ера	Ipc	Epc	Ipa/	Epa/	Cathodic	Anodic
mM	μA	mV	μA	mV	Ірс	Epc	rate	rate
0.001	284.1	107.2	472.5	1066	0.601	958.8	0.163	0.098
0.002	361	127.3	459	1111	0.786	983.7	0.158	0.124
0.003	361.2	120.7	462	1116	0.781	995.3	0.159	0.124
0.004	409	142.5	451	1235	0.906	1092.5	0.155	0.141
0.005	419.2	142.9	448	1159	0.935	1016.1	0.154	0.144
0.006	425	139.4	509	1080	0.834	31.4	0.175	0.146
0.007	438.6	185.4	415	1150	1.056	70.4	0.143	0.151
0.008	450	185	446	1121	1.008	936	0.154	0.155
0.009	462	180	450	1220	1.026	58	0.155	0.159
0.01	481	175	444	1250	1.083	50	0.153	0.166

Table 4. Different concentrations (0.001-0.01 mM) of sodium saccharin in 1 M Na₂SO₃, at cathodic and anodic current peaks, by CNT/GCE.

The expression for the reaction rate depends upon the electrode area, A $[cm^2]$, as shown in Table 4¹⁷,

Rate
$$=\frac{I}{nAF}$$
 (3)

where I is the current, F is the number of faradays, A is the electrode area, and n is the number of transferred electrons (n = 1).

Figs. 7 and 8 show that the cathodic and anodic rate reactions of sodium saccharin were proportional to the different concentrations (0.001-0.01 mM) in both oxidation and reduction processes.

The redox rate reaction increased against increasing concentrations of sodium saccharine in the electrolyte¹⁸.

Table 4 shows the reaction rate at anodic and cathodic electrodes, with different concentrations (0.001-0.01 mM) of sodium saccharin in 1 M Na₂SO₃. The relationship between the reaction rate and the sodium saccharin concentration on the anodic and cathodic electrodes is different in terms of properties, as shown in Figs. 7 and 8, respectively. The oxidation reaction rate at the anodic electrode increased against an increasing concentration of sodium saccharin in the electrolyte, which depended on a lower reduction rate at the cathodic electrode, as shown in the following equation:





Figure 7. Relationship between the anodic reaction rate and different concentrations of sodium saccharin in 1 M Na₂SO₃, at CNT/GCE.



Figure 8. Relationship between the cathodic reaction rate and different concentrations of sodium saccharin in 1 M Na₂SO₃, at CNT/GCE.

Effect of varying the scan rate

The voltammograms of 0.01 mM sodium saccharin in 1 M Na₂SO₃ were studied at different scan rates, which express that an increased scan rate followed increased redox current peaks, as shown in Figs. 9 and 10. However, the anodic current peak shifted towards higher potentials, and the cathodic current peak shifted to lower potentials, as shown in Fig. 11. In a slow voltage scan, the diffusion layer grows much further from the electrode, in comparison to a fast scan. As a result, the flux to the electrode surface is much lower at slow scan rates than at faster ones. As the current is proportional to the flux towards the electrode, the current intensity becomes lower at slow scan rates, and higher at high scan rates¹⁹.



Figure 9. Plot log of I_{pa} against Log V (scan rate) for 1 mM sodium saccharin in 1 M Na₂SO₃ and 0.1 M borate buffer, at pH 3, using CNT/GCE.



Figure 10. Plot log of Ipc against Log V (scan rate) for 1 mM sodium saccharin in 1 M Na₂SO₃ and 0.1 M borate buffer, at pH 3, using CNT/GCE.

Scan Rate Vs ⁻¹	Ірс µА	Epc mV	Іра µА	Epa mV	Epa-Epc mV	Ipa/Ipc	D _f , pc m ² /s x 10 ⁻⁴	D _f , pa m ² /s x 10 ⁻⁴
0.01	655.3	830.4	146.4	14.06	-816.34	0.223	0.659	0.032
0.02	832.2	917.5	249.5	47.68	-869.82	0.299	0.531	0.047
0.03	936.1	952.1	454.9	92.97	-859.13	0.485	0.448	0.105
0.04	1046	981.1	545.7	111.7	-869.4	0.521	0.420	0.114
0.05	1123	972.4	596.3	120.3	-852.1	0.530	0.387	0.109
0.06	1193	981.3	650.9	139	-842.3	0.545	0.364	0.108
0.07	1267	997.7	698.7	143.4	-854.3	0.551	0.352	0.107
0.08	1314	1030	799	167.6	-862.4	0.608	0.331	0.122
0.09	1487	1012	769.2	167.4	-844.6	0.517	0.377	0.100
0.1	1491	1020	1091	203.4	-816.6	0.731	0.341	0.182

Table 5. Redox current, potential, peak potential separation, peak current ratio and diffusion coefficient (D_f) of sodium saccharin, at different scan rates, on CNT/GCE.

A reasonably linear dependence of sodium saccharin's reduction and oxidation currents on different scan rates was described by the equations y = 0.3551x + 3.5181 (R² = 0.9903), and y = 0.8018x + 3.7904 (R² = 0.9681), which are displayed in Figs. 9 and 10. It was found that the ratio of anodic to cathodic current peaks increased with increasing scan rates.

The system in the redox ratio of the voltammograms is recorded in Table 5.

The diffusion coefficient values of the redox process of sodium saccharin at different scan rates were calculated from Randles-Seveik equation, as described in Table 5. It was found that the diffusion coefficient values were close to the values for the pollutant materials in the environment²².



Figure 11. Cyclic voltammogram of sodium saccharin in pH=3 at different scan rates (0.01 and 0.1 mV sec⁻¹), on CNT/GCE.

Fig. 11 shows the cyclic voltammograms of sodium saccharin at different scan rates (0.01 and 0.1mVsec^{-1}), on CNT/GCE. Randles-Seveik equation^{20,21} describes a reversible redox couple of the current peaks.

$$I_{p} = (2.69 \text{ x } 10^{5}) \text{ n}^{3/2} \text{ AC } D_{f}^{1/2} \text{ v}^{1/2}$$
(3)

where Ip is the current, n is the moles number of electrons transferred into the reaction, A is the electrode area, D_f is the diffusion coefficient, and V is the scan rate of the applied potential.

Reliability and stability of modified electrode

The potential cycling of the oxidation-reduction current was carried out during cyclic voltammetry for the modified CNT/GCE working electrode in sodium saccharine with Na₂SO₃, at a scan rate of 100 mVsec⁻¹ (Tables 6 and 7). Table 6 illustrates the reliability of the anodic current peak (I_{pa}) current, and the relative standard deviation (RSD), which is $\pm 1.95\%$.

Table 6. Reliability of CNT/GCE as working electrode, at a scan rate of 100 mVsec⁻¹, for the anodic current peak of 0.01 mM sodium saccharin in 1 M Na₂SO₃, at nine times cycles.

Number	Іра	Mean	RSD
1	180.6	182.96	±2.073%
2	183.4		
3	184.5		
4	183.8		
5	183.8		
6	184.9		
7	186.8		
8	189.9		
9	176.2		
10	175.7		

Table 7. Reliability of CNT/GCE as working electrode, at a scan rate of 100 mVsec⁻¹, for the cathodic current peak of 0.01 mM sodium saccharin in 1 M Na₂SO₃, at ten times cycles.

Number	Ірс	Mean	RSD
1	341.2	335.31	±0.739%
2	336.4		
3	333.8		
4	332.9		
5	334.5		
6	336.1		
7	336.4		
8	335.3		
9	333.7		
10	333.8		



Figure 12. Cyclic voltammogram of redox current peaks of 0.01 mM sodium saccharin in 1 M Na₂SO₃ (pH=3), at ten* times cycles, on CNT/GCE, at a scan rate of 100 mV sec⁻¹ versus Ag/AgCl.

Fig. 12 shows the redox current peaks CV of 0.01 mM sodium saccharine with 1 M Na_2SO_3 at ten times cycles, which revealed a good stability of the modified GCE CV, by overlapping of the voltammogram lines.

Conclusion

The results of the research showed that the oxidation – reduction current peaks of sodium saccharine in different electrolytes were enhanced by a nano-sensor (CNT/GCE) in electrochemical analysis, using the cyclic voltammetric method. The oxidation – reduction current peaks of the sodium saccharin in Na₂SO₃, at the potentials of 102.9 and -563 mV, respectively, were studied at different concentrations, pH values, and scan rates. The results indicate that sodium saccharin has high oxidation-reduction current peaks which were enhanced at acidic pH (=5) in Na₂SO₃ as an electrolyte solution. Also, the redox current peaks of sodium saccharin were studied at different scan rates, which determined the current's ratio (Ipa/Ipc \approx 1) and the peak potential separation (Epa – Epc \approx 100 mV). This demonstrates that the redox process of saccharin in an aqueous electrolyte solution is reversible, and that there was a heterogeneous electron transfer into the reaction. The diffusion coefficient value for sodium saccharin on the CNT/GCE surface had an average value of 0.421×10^{-4} m²/sec for the cathodic electrode, and of 0.103×10^{-4} for the anodic electrode. Another factor, the redox reaction rates, was determined, to study the relationship with the different concentrations of sodium saccharin in the electrolyte solution, which gave different properties of the oxidation current peak from those of the reduction current peak.

References

- 1. Radhi MM, Abdul-sahib AM, J Biochem Intern. 2016;3(1):1-8.
- 2. Radhi MM, Abdul-Amir YK, Khalaf MS. Electrochemical Effect of Acetylsalicylic Acid (Aspirin) in Present of Each Ascorbic Acid (AA) and Folic Acid (FA) in Normal Saline and Human Blood Samples. AASCIT Communications. 2016;3(3):152-9.
- 3. Radhi MM, Albakry AA, Jassim AM, et al. Electrochemical Study of Pb(II) in Present of Each Ascorbic Acid, Glucose, Urea and Uric Acid Using Blood Medium as an Electrolyte. Nano Biomed Eng. 2016;8(1):9-15. Doi: https://doi.org/10.5101/nbe.v8i1.p9-15
- 4. Radhi MM, Abdullah HN, Al-Asadi SA, et al. Electrochemical oxidation effect of ascorbic acid on mercury ions in blood sample using cyclic voltammetry. Int J Ind Chem. 2015;6:311-6. Doi: https://doi.org/10.1007/s40090-015-0053-9
- 5. Radhi MM, Khalaf MS, Ali ZO, et al. Voltammetric Analysis of Zn (II) in Present of Each Ascorbic Acid (AA) and Folic Acid (FA) in Human Blood Samples. AASCIT Communications. 2016;3(1):11-6.
- 6. Zubrick JW. The Organic Chem Lab Survival Manual. 4th edition. New York: Wiley & Sons, Inc; 1997.

- Islam GJ, Naseem HM, Mamun MA, et al. Investigations on the redox behaviour of manganese in manganese(II)–saccharin and manganese(II)– saccharin–1,10-phenanthroline complexes. J Saudi Chem Soc. 2009;13(2):177-83. Doi: https://doi.org/10.1016/j.jscs.2009.05.002
- Ivanova V, Herman CB. Adsorption of saccharin on Au(1 1 1) single crystals. J Electroanal Chem. 2003;552:45-51. Doi: https://doi.org/10.1016/S0022-0728(03)00211-0
- Tanvir T, Hossain Md, Mamun, et al. Preparation and characterization of iron(III) complex of saccharin. J Bangladesh Acad Sci, 2013; 37:2: 195-203. DOI: https://doi.org/10.3329/jbas.v37i2.17560
- Chen Q, Mou S, Liu K, et al. Separation and determination of four artificial sweeteners and citric acid by high-performance anion-exchange chromatography. J Chromatography A. 1997;771(1-2):135-143.Doi: https://doi.org/10.1016/S0021-9673(97)00067-8
- 11. Çakır S, Biçerb E. Synthesis, spectroscopic and electrochemical characteristics of a novel Schiff-base from saccharin and tryptophan. J Iran Chem Soc. 2010;7(2)394-404. Doi: https://doi.org/10.1007/BF03246025
- 12. Rahman MT, Hossain E, Ehsan M. Spectrophotometric and cyclic voltammetric study of interaction of Fe(III) with vitamin B3 and vitamin B6. J Bangladesh Acad Sci. 2014;38(2):143-53. Doi: https://doi.org/10.3329/jbas.v38i2.21339
- Masdek NR, Alfantazi AM. An EQCM study on the influence of saccharin on the corrosion properties of nanostructured cobalt and cobalt-iron alloy coatings. J Solid State Electrochem. 2014;18(6):1701-16. Doi: https://doi.org/10.1007/s10008-014-2417-z
- 14. Scholz F, Lange B. Abrasive stripping voltammetry an electrochemical solid state spectroscopy of wide applicability. TrAC Trends Anal Chem.1992;11(10):359-67. Doi: https://doi.org/10.1016/0165-9936(92)80025-2
- 15. Tan WT, Ng GK, Bond AM. J. Chem. 2000;2(2):34-42.
- 16. Tan WT, Farhan Y, Zulkarnain Z. Sensors Transducers J. 2009;104(5):119-27.
- 17. Sutter EA, Sutter PW. J Am Chem Soc. 2014;136(48):16865-70. https://doi.org/10.1021/ja508279v
- 18. Bard AJ, Faulkner LR. Electrochemical Methods: Fundamentals and Applications. 2nd Ed. New York: Wiley; 2001.
- Farhana H, Rahman MS, Etmina A, et al. A Cyclic Voltammetric Study of the Redox Reaction of Cu(II) in Presence of Ascorbic Acid in Different pH Media. Dhaka Univ J Sci. 2013;61(2):161-166. Doi: https://doi.org/10.3329/dujs.v61i2.17064
- 20. Zanello P. Inorganic Electrochemistry: Theory, Practice and Application. London: The Royal Society of Chemistry; 2003.
- 21. Crouch SR, Skoog DA. Principles of instrumental analysis. UK: Cengage Learning; 2006.
- 22. Cussler EL. Diffusion: Mass Transfer in Fluid Systems. 2nd edition. New York: Cambridge University Press; 1997.